

CGS 27830, A POTENT NONPEPTIDE ENDOTHELIN RECEPTOR ANTAGONIST

Ben Mugrage*, John Moliterni, Leslie Robinson, Randy L. Webb, Suraj S. Shetty,
Kenneth E. Lipson, Michael H. Chin, Robert Neale and Catherine Cioffi
Ciba-Geigy Corp., Pharmaceuticals Div., 556 Morris Ave., Summit, NJ 07901

(Received in USA 13 July 1993; accepted 13 August 1993)

Abstract: Treatment of the dihydropyridine carboxylic acid **1** with EDCI produced a diastereomeric mixture of chemically stable anhydrides. The meso-isomer, **CGS 27830**, was isolated by crystallization and found to be a potent (16 nM) endothelin receptor antagonist for the ET_A receptor subtype.

We wish to report the synthesis and biological activity of a potent nonpeptide endothelin antagonist derived from the dihydropyridine class of calcium ion modulators.¹ The symmetric anhydride (**CGS 27830**) was found to have rather poor potency in inhibiting the binding of [³H]PN200-110 to L-type calcium channels (IC₅₀ = 592 ± 94 nM) in rat cortex membranes.² However, **CGS 27830** was found to potently inhibit the binding of [¹²⁵I]ET-1 to the ET_A receptor subtype (IC₅₀ = 15.9 ± 1.3 nM) in porcine thoracic aorta membranes.^{3,12} Using rat cerebellum membranes, **CGS 27830** was also able to compete for binding to the ET_B receptor subtype, but with considerably less affinity (IC₅₀ = 295 ± 19 nM).

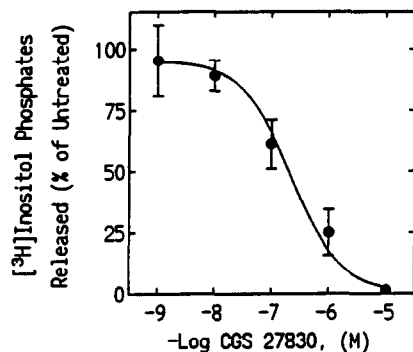


Figure 1. Ability of CGS 27830 to inhibit [³H]inositol phosphate accumulation in A7r5 cells. Intact cells were preincubated with CGS 27830 for 30 min and then challenged with 5 nM ET-1 for an additional 30 min. Values are shown as the mean ± SEM from 3 independent experiments.

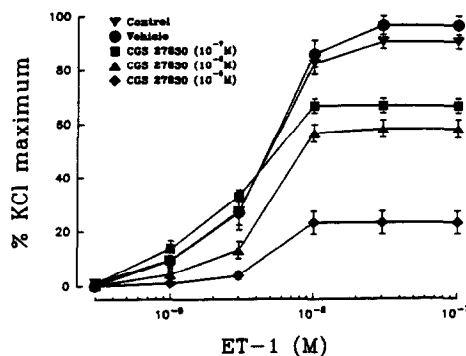


Figure 2. Effect of CGS 27830 on contractions evoked by ET-1 in isolated rabbit aortic strips. The tissues were equilibrated with CGS 27830 for 30 min before exposure to cumulative concentrations of ET-1. All changes in force are expressed as percentage of maximal response to KCl and expressed as the mean ± SEM.

Further biological profiling showed that CGS 27830 inhibited ET-1-induced phosphoinositide (PI) turnover ($EC_{50} = 143 \pm 58$ nM, $n = 3$) in A7r5 smooth muscle cells (fig. 1), while having no effect on basal PI turnover.⁴ Since the ET_A receptor predominates in this cell line, these data demonstrate the potency of CGS 27830 as an ET_A antagonist. A 30 min pretreatment with CGS 27830 (0.1 - 10 μ M) produced a dose dependent, insurmountable inhibition of the ET-1 induced contractile response in isolated rabbit aorta. The maximal response to ET-1 was attenuated by 33, 42 and 77% at 0.1, 1 and 10 μ M, respectively (fig. 2). Similar pretreatment with CGS 27830 (1 μ M) did not alter the concentration curves for phenylephrine or KCl suggesting that the antagonist action of CGS 27830 is specific for ET-1. A 5 min pretreatment with CGS 27830 (10 mg/kg, i.v.) abolished the pressor response and attenuated the depressor response to ET-1 (0.25 nmol/kg i.v.) in the conscious rat (fig. 3). Unfortunately, an assessment of the duration of action of CGS 27830 revealed a relatively short half-life ($T_{1/2} < 60$ min) as shown in fig. 4.

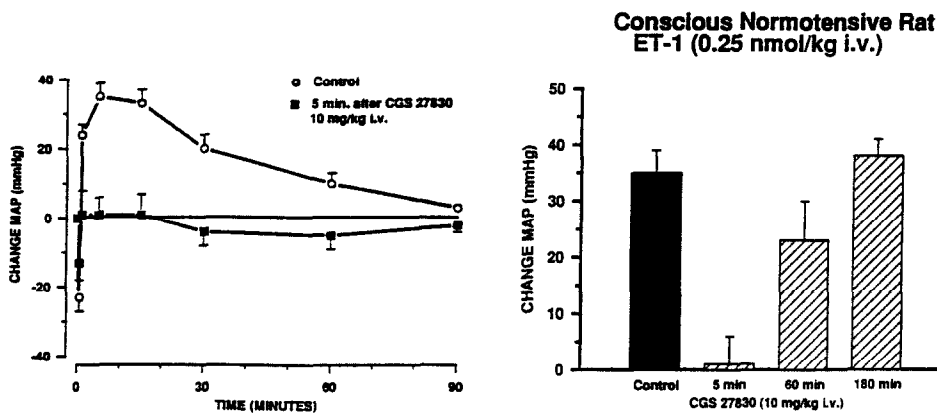
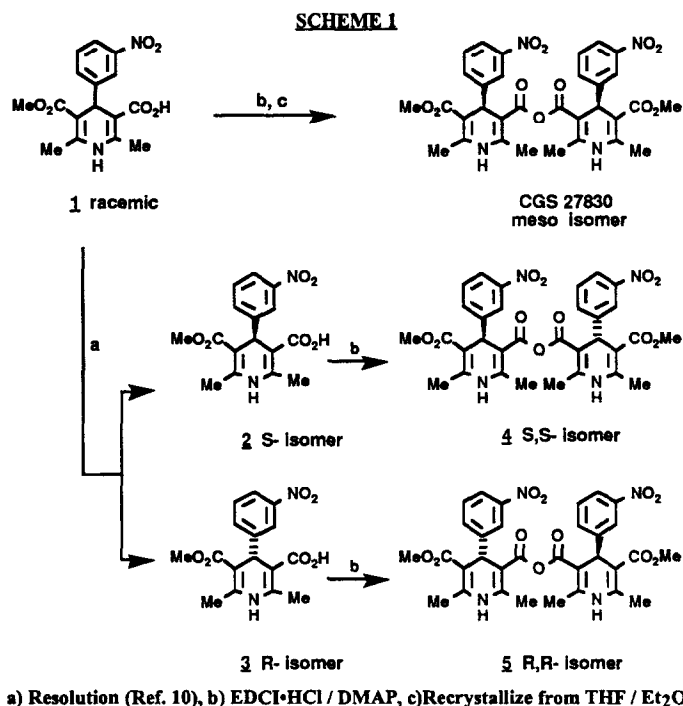


Figure 3. Effect of CGS 27830 on ET-1-induced changes in mean arterial blood pressure in the conscious Wistar Kyoto rat. CGS 27830 (10 mg/kg, i.v.) or vehicle were administered 5 min prior to ET-1 (0.25 nmol/kg, i.v.).

Figure 4. Effect of CGS 27830 on ET-1-induced increases in blood pressure. CGS 27830 was given either 5, 60 or 180 min prior to an ET-1 challenge (hatched bars). Changes in blood pressure evoked by ET-1 in the absence of the antagonist are also shown (solid bar).

The other possible diastereomeric anhydrides were also examined for ability to antagonize ET-1 binding. The R,R-isomer (5) was found to only moderately inhibit binding (IC_{50} ; $ET_A = 422 \pm 56$ nM, $ET_B = 2.7 \pm .5$ μ M), while the S,S-isomer (4) was found to be inactive at both endothelin receptor subtypes.



A diastereomeric mixture of anhydrides was produced, as shown in Scheme 1, by treatment of the known dihydropyridine carboxylic acid⁵ with a dehydrating agent. The most efficient method was the utilization of 1-ethyl-3-(3-dimethylamino-propyl)carbodiimide hydrochloride (EDCI·HCl) with a catalytic amount of dimethylaminopyridine (DMAP) in CH₂Cl₂, at room temperature for 1h.⁶ Other dehydrating conditions such as oxalyl chloride / pyridine or diethylazodicarboxylate (DEAD) with triphenylphosphine also gave the mixture of diastereomeric anhydrides, although in much lower yields.⁷ The anhydride proved remarkably stable and could be washed with water or 1N K₂CO₃ solution without appreciable hydrolysis. Another reflection of the surprising stability of the anhydride was that flash chromatography did not result in decomposition. CGS 27830 was separated from the mixture of stereoisomers, as a light yellow solid, by careful crystallization from THF / Et₂O. A second crystallization from THF / Et₂O produced crystals suitable for x-ray structure determination (fig. 5).⁸

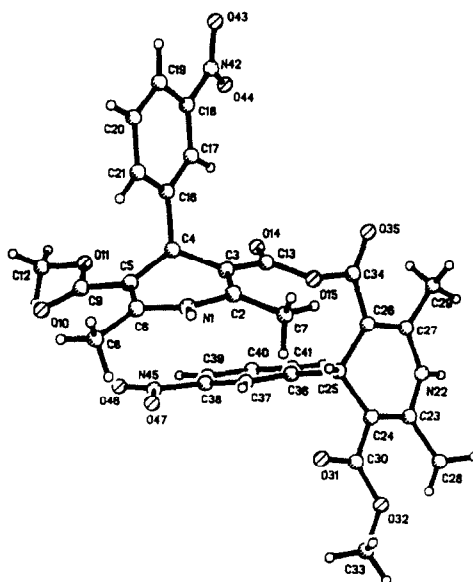


Figure 5. The x-ray structure of CGS 27830

The R,R and S,S-stereoisomers⁹ (4, 5) were produced from the corresponding optically active dihydropyridine carboxylic acids¹⁰ (2, 3) in a similar fashion. The des-nitro, *m*-chloro and *m*-methoxy anhydride analogs were also tested, as diastereomeric mixtures, and found to have receptor binding activities similar to the *m*-nitro compound.

We have shown CGS 27830 to be a potent and selective endothelin receptor antagonist. Results from receptor binding studies indicate that CGS 27830 displays more than an 18-fold selectivity for the ET_A receptor subtype. However, results from in vitro and in vivo studies suggest some functional ET_B antagonist activity as well. Given the insurmountable nature of the receptor antagonism and the potential for anhydrides to act as acylating agents, covalent modification of the receptor by CGS 27830 is a distinct possibility. Experiments to determine the mode of action of CGS 27830 are ongoing. Nevertheless, the potent and readily synthesized receptor antagonist CGS 27830 represents a valuable pharmacologic tool for the investigation of the role of ET-1 in models of vascular disease states.¹¹

Acknowledgment: Dr. Frank H. Clarke is gratefully thanked for the x-ray structure determination studies. Drs. Herman Rodriguez and Gary Ksander are thanked for valuable discussions.

References and Notes

1. F. Bossert, H. Meyer and E. Wehinger; *Angew. Chem. Int. Ed. Eng.*, **20**, 762, (1981).
2. The binding of [³H]PN200-110 to rat cortex membranes was performed by the method of Lee *et al.*: Lee, H.R., Roeske, W.R., and Yamamura, H.I., *Life Sci*, **35**, 721 (1984).
3. The binding of [¹²⁵I]ET-1 to porcine thoracic aorta membranes (ET_A), or rat cerebellum membranes (ET_B) was performed as previously described for rat lung membranes: Cioffi, C., Neale Jr., R.F., Jackson, R.H., and Sills, M.A., *J. Pharmacol. Exp. Ther.*, **262**, 611 (1992).
4. The release of [³H]inositol phosphates from [³H]inositol-labeled A7r5 cells was measured as previously described: Cioffi, C. and Garay, J., *J. Cardiovas. Pharmacol.*, in Press (1993).
5. E. Wehinger and F. Bossert; US Pat. 4,285,955, (1979).
6. Preparation of **CGS 27830**: 1,4-Dihydro-5-methoxycarbonyl-2,6-dimethyl-4-(3-nitrophenyl)-3-pyridine-carboxylic acid (**1**) (21.4 g, 0.065 mol) and 1-ethyl-3-(3-dimethylaminopropyl)-carbodiimide hydrochloride (23.0 g, 0.120 mol) were suspended in dry CH₂Cl₂ (300 ml) and treated with a catalytic amount (25 mg) of 4-dimethylaminopyridine. The reaction was stirred, at room temperature, under an N₂ atmosphere, until a clear yellow solution was obtained (~2h). Tlc (SiO₂ / Et₂O) indicated the reaction was complete. The solution was transferred to a separatory funnel and washed sequentially with water (200 ml), 1N K₂CO₃ (100 ml) and brine (100 ml). The organic layer was dried (MgSO₄) and concentrated to a yellow foam. The crude product was purified by passing through a short plug of flash silica gel eluted with 1:1 Et₂O / EtOAc followed by EtOAc to produce, after evaporation, a 2:1 mixture of diastereomers favoring the meso-isomer. The isomeric mixture was then dissolved in THF (600 ml) and Et₂O added until the solution became slightly cloudy. The solution was kept at 4°C overnight to give **CGS 27830** (6.0 g) as a light yellow solid (mp 223-4° dec) which was isolated by decantation.
¹H-NMR (DMSO-d₆) 9.38, s, 2H; 8.03, d, 2H, J = 8.3; 7.70, s, 2H; 7.46-7.57, m, 4H; 4.58, s, 2H; 3.50, s, 6H; 2.26, s, 6H; 2.19, s, 6H., ¹H-NMR (CDCl₃) 8.00, m, 2H; 7.85, m, 2H; 7.33-7.50, m, 4H; 5.95, s, 2H; 4.72, s, 2H; 3.60, 6H; 2.30, s, 6H; 2.26, s, 6H. IR (CH₂Cl₂) 3350, 1751, 1701, 1652, 1609, 1530, 1472, 1435, 1351, 1225, 1185, 1112, 1061, 950 cm⁻¹. Calc. for C₃₂H₃₀N₄O₁₁: 59.44 %C, 4.68 %H, 8.67 %N; Found: 59.25 %C, 4.50 %H, 8.52%N.

7. When the dihydropyridine carboxylic acid **1** and EDCI·HCl are stirred in the presence of 1-hydroxybenzotriazole, a stable HOBT adduct is formed that can be isolated and flash-chromatographed.
8. The refined coordinates, bond distances, bond angles and torsion angles with esd's for the crystal structure of **CGS 27830** have been submitted for deposition at the Cambridge Crystallographic Data Centre.
9. Analytical data for **4**: ¹H-NMR (CDCl₃) 8.05, m, 2H; 7.98, m, 2H; 7.60, d, 2H, J = 8.1 Hz; 7.33, t, 2H, J = 8.1 Hz; 6.05, s, 2H; 5.12, s, 2H; 3.70, s, 6H; 2.37, s, 6H; 2.23, s, 6H. Calc. for C₃₂H₃₀N₄O₁₁: 59.44 %C, 4.68 %H, 8.67 %N; Found: 59.13 %C, 4.50 %H, 8.33 %N. [α]_D (c 0.54, CHCl₃) = -269°. Analytical data for **5**: ¹H-NMR (CDCl₃) 8.02, m, 2H; 7.95, m, 2H; 7.59, d, 2H, J = 8.1 Hz; 7.31, t, 2H, J = 8.1 Hz; 6.19, s, 2H; 5.10, s, 2H; 3.70, s, 6H; 2.34, s, 6H; 2.20, s, 6H. Calc. for C₃₂H₃₀N₄O₁₁: 59.44 %C, 4.68 %H, 8.67 %N; Found: 59.63 %C, 4.53 %H, 8.46 %N. [α]_D (c 0.81, CHCl₃) = +329°.
10. A. Ashimori, T. Uchida, Y. Ohtaki, M. Tanaka, K. Ohe, C. Fukaya, M. Watanabe, M. Kagitani and K. Yokoyama; *Chem. Pharm. Bull.*, **39**, 108, (1991).
11. a) Nguyen, P., Parent, A., Deng, L., Flückinger, J., Thibault, G., Schriffin, E., *Hypertension (Supp. II)*, **19**, II98 (1992). b) Deng, L. and Schriffin, E., *Am. J. Physiol. (Heart Physiol 31)*, **262**, H1782 (1992). c) Clozel, M., *J. Hypertens.*, **7**, 913 (1989) d) Schriffin, E., Deng, L., Larochelle, P., *J. Hypertens.*, **10**, 437 (1992). e) Shichiri, M., Hirata, Y., Emori, T., Ohata, K., Ogura, M., Inoue, A., Marumo, F., *Hypertension*, **15**, 493 (1990). f) Firth, J., Ratcliffe, P., Raine, A., Ledingham, J., *The Lancet*, 1179 (1988). g) Kon, V., Yoshioka, T., Fofo, A., Ichikawa, I., *J. Clin. Invest.*, **83**, 1762 (1989). h) Papadopoulos, S., Gilbert, L., Webb, R., D'Amato, C., *Neurosurgery*, **26**, 810 (1990).
12. Using the cyclic pentapeptide endothelin antagonist, BQ-123, as a reference in our assays the following results were obtained: ET_A receptor binding, IC₅₀ = 22 nM; PI-turnover IC₅₀ = 87 nM; Isolated rabbit aorta BQ-123 (1 μM) inhibited 28% of the contraction caused by ET-1 (0.1 μM); In Vivo, in conscious rats, a 5 min. pretreatment with BQ-123 (10 mg/kg, i.v.) inhibited 57% of the pressor response caused by ET-1 (0.25 nmol/kg, i.v.).
13. Nonpeptide endothelin receptor antagonists has been reported previously see: a) S-i. Mihara and M. Fujimoto, *Eur. J. Pharmacol., Mol. Pharmacol Sect.*, **246**, 33 (1993). b) M. Fujimoto, S-i. Mihara, M. Ueda, M. Nakamura and K-s Sakurai, *FEBS Lett.*, **305**, 41 (1992). c) R. Cousins *et al.*, World Pat. 9308799 (1993). d) K. Burri *et al.*, Eur. Pat. 510,526A (1992). e) O. Nobutaka *et al.*, Eur. Pat. 405,421 (1991).